

# **Borrowing information from relevant microarray studies for sample classification using weighted partial least squares**

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# Outline

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  - Partial least squares (PLS)
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# Introduction

- Traditional medical diagnosis/classification method is very subjective
  - Based on morphological characteristics, pathological features
  - Depends on highly trained pathologists

**Limitation:** Hard to diagnose disease subtypes that are morphologically similar but follow different clinical courses.

- New classification method is objective
  - Based on microarray gene expression data.
  - Can be highly accurate.

**Potentials:** diagnose disease subtypes; predict clinical outcomes...

# Example: Two-class microarray

- Notations:

	Class 1			Class 2		
	1	...	$n_1$	$n_1 + 1$	...	$n_1 + n_2 = n$
gene 1	$X_{1,1}$	...	$X_{1,n_1}$	$X_{1,n_1+1}$	...	$X_{1,n}$
gene 2	$X_{2,1}$	...	$X_{2,n_1}$	$X_{2,n_1+1}$	...	$X_{2,n}$
⋮	⋮	⋮	⋮	⋮	⋮	⋮
gene $p$	$X_{p,1}$	...	$X_{p,n_1}$	$X_{p,n_1+1}$	...	$X_{p,n}$

- Outcome  $Y = (y_1, y_2, \dots, y_n)'$ .

- Covariates  $X_i = (x_{i1}, x_{i2}, \dots, x_{in})'$ ,  $i = 1, \dots, p$ .  
Covariates are often standardized  $var(x_i) = 1$ .

- A special feature of microarray data:

Small  $n$ , large  $p$

- A simple prediction problem:
  - Our goal is to predict  $Y$  from  $X_1, X_2, \dots, X_p$  by a linear model.
  - Especially interested in problems where  $p \gg n$ .
- Many new methods have appeared
  - Weighted voting, Compound covariate,...
  - Penalized regression: Shrunken centroids, LASSO,...
  - Machine learning: SVM, Bagging/boosting trees,...

# Penalized Partial Least Squares

- Partial Least Squares (PLS)
  - Particularly suited for constructing linear models when there are more variables than observations.
  - Robust to the collinearity between covariates.
  - Suited for fitting linear models with microarray data.
- Penalized Partial Least Squares (PPLS)
  - A penalized regression method built on the framework of PLS.

Ref: **Huang, X.** and **Pan, W.** (2003). Linear regression and two-class classification with gene expression data. *Bioinformatics* **19**, 2072-2078.

# Application to the Minnesota data: PPLS

## ● Minnesota data

- Oligonucleotide microarray data obtained by Hall et al. (2003) in a heart failure study conducted at the Medical School of UMN.
- Contain 30 samples: 10 ischemic, 7 ischemic with acute MI and 13 idiopathic.
- Affy HG-U133A chips: Contain 22,283 genes.
- Initially processed in MAS 5.0.

**Goal:** Distinguish between the ischemic and the idiopathic etiology classes.

# Initial gene ranking

- Given gene  $i$

$$F_i = \frac{MS_{class}}{MS_{error}} = \frac{\left(\sum_{c=1}^C n_c (\bar{x}_{i_c} - \bar{x}_i)^2\right) / (C - 1)}{\left(\sum_{c=1}^C \sum_{j \in c} (x_{ij} - \bar{x}_{i_c})^2\right) / (n - C)}.$$

$x_{ij}$ : gene expression intensity of gene  $i$  and sample  $j$ .

$n_c$ : number of samples in class  $c$ ,  $n = \sum_{c=1}^C n_c$ : total sample size.

$C$ : number of classes.

$\bar{x}_{i_c}$ : mean gene expression of class  $c$ .

$\bar{x}_i$ : overall mean gene expression.

- Genes with larger F-statistics -> higher rank.

# Experiment: Minnesota data

- LOOCV error (Isch vs Idio,  $n = 23$ )

# of top genes	PPLS	SC	LASSO
50	10	5	6
100	7	5	7
200	9	7	11
800	6	8	4
1600	5	8	8
9600	9	8	7
16000	8	7	8
22283	9	5	7

# Application to the PGA data: PPLS

- PGA data
  - Oligonucleotide microarray data obtained in a heart failure study conducted at the PGA Medical School.
  - Contain 36 samples: 11 normal, 11 ischemic, and 14 idiopathic.
  - Affy HG-U133 plus 2 chips: Contain  $\sim 54,000$  genes.
  - Initially processed in MAS 5.0.

**Goal:** Distinguish between the ischemic and the idiopathic etiology classes.

# Experiment: PGA data

- LOOCV error (Isch vs Idio,  $n = 25$ )

# of top genes	PPLS	SC	LASSO
50	3	2	2
100	1	2	1
200	1	2	1
800	1	1	3
1600	1	1	2
9600	1	1	1
16000	1	1	1
22277	1	1	1

# A summary on the above experiments

- The LOOCV misclassification error
  - ranges from 5 to 11 for the Minnesota data (23 samples).
  - ranges from 1 to 3 for the PGA data (25 samples).
- These highlight some existing differences underlying the two datasets.
- **A question: Is there any signal/predictive information in the data?**

# Permutation test: Minnesota data

- LOOCV error (Isch vs Idio,  $n = 23$ )

# of top genes	Original data		Permutated data				
	CV errors	P-value	0%	25%	50%	75%	100%
50	5	.00	5	10	11	12.75	19
100	5	.00	5	10	11	12	19
400	7	.06	4	9	11	13	17
1600	8	.08	5	9.25	11.5	13	17
6400	9	.12	6	10	11	13	21

# Borrow information from relevant studies

- To increase the statistical power, borrow information from other relevant studies.
- A key difference from meta-analysis:
  - Not assuming current study shares a common set of parameters with other studies.
- Example: Identifying genes associated with ventilator-associated lung injury (VALI) based on a human study.
  - Meta-analysis: Only interested in the genes associated with VALI which are conserved across the species over the evolutionary history (Grigoryev et al. 2004).
  - Our analysis: Interested in inference on a set of parameters specific for humans.

# Combining Minnesota and PGA data

- Classification with the combined data.
  - Goal: Distinguish etiologies of heart failure for Minnesota patients while treat the PGA data as secondary.
  - Problem: Unobserved differences in patient characteristics.
  - Solution: Treat samples in different studies unequally, e.g, assign different weights.
- Combining Minnesota data and PGA data.
  - Technically easy: all probe sets present on U133A chip are identically replicated on U133 Plus 2 chip.
  - Data mapped by probe set ID (6 could not be found).

# Notations

- Given  $X$ , to predict  $Y$  with a linear model

$F(X, \mathbf{a}) = a_0 + \sum_{i=1}^p a_i x_i$ , the goal is to minimize the expected loss (risk):

$$R(\mathbf{a}) = E_Y L(Y, F(X, \mathbf{a})).$$

- $L(Y, F(X, \mathbf{a}))$ : loss criterion.
- An empirical estimate of the expected loss:

$$\hat{R}(\mathbf{a}) = \frac{1}{n} \sum_{j=1}^n L(y_j, a_0 + \sum_{i=1}^p a_i x_{ij}).$$

- The optimal values of  $\mathbf{a}$ :

$$\hat{\mathbf{a}} = \arg \min_{\mathbf{a}} \frac{1}{n} \sum_{j=1}^n L(y_j, a_0 + \sum_{i=1}^p a_i x_{ij}).$$

# Partial Least Squares (PLS)

- Conjugate gradient procedure under squared error loss:

$$\begin{aligned}\hat{\mathbf{a}}_{k+1} &= \hat{\mathbf{a}}_k + \rho_k \mathbf{s}_k \\ \mathbf{s}_k &= \mathbf{g}_k + \frac{\mathbf{g}_k^T \mathbf{g}_k}{\mathbf{g}_{k-1}^T \mathbf{g}_{k-1}} \mathbf{s}_{k-1}\end{aligned}$$

- $\mathbf{g}_k$ : negative gradient at  $\hat{\mathbf{a}}_k$

$$\mathbf{g}_k = -\frac{\partial}{\partial \mathbf{a}} \hat{R}(\mathbf{a}) \Big|_{\mathbf{a}=\hat{\mathbf{a}}_k}$$

- $\rho_k$ : step size

$$\rho_k = \operatorname{argmin}_{\rho} \hat{R}(\hat{\mathbf{a}}_k + \rho \mathbf{s}_k)$$

- $k$ : number of PLS components.

- Squared error loss:  $L(Y, F(X, \mathbf{a})) = (Y - F(X, \mathbf{a}))^2 / 2$ .

# Propose: Weighted PLS

- Expected loss estimated by a weighted average loss:

$$\hat{R}_w(\mathbf{a}) = \sum_{j=1}^n w_j L(y_j, a_0 + \sum_{i=1}^p a_i x_{ij}).$$

- Conjugate gradient procedure under squared error loss:

$$\mathbf{g}_k = -\frac{\partial}{\partial \mathbf{a}} \hat{R}_w(\mathbf{a}) \Big|_{\mathbf{a}=\hat{\mathbf{a}}_k} = Z^T W (Y - Z\hat{\mathbf{a}}_k)$$

$$\rho_k = \operatorname{argmin}_{\rho} \hat{R}_w(\hat{\mathbf{a}}_k + \rho \mathbf{s}_k) = \begin{cases} 1 & \text{if } Z\mathbf{s}_k = 0 \\ \frac{(Z\mathbf{s}_k)^T W (Y - Z\hat{\mathbf{a}}_k)}{(Z\mathbf{s}_k)^T W (Z\mathbf{s}_k)} & \text{if } Z\mathbf{s}_k \neq 0 \end{cases}$$

- $W = \operatorname{diag}(w_1, \dots, w_n)$ : diagonal matrix with weights,  $\sum_{j=1}^n w_j = 1$ .

- $Z = (\mathbf{1} \ X_1 \ X_2 \ \dots \ X_p)_{n \times (p+1)}$ : covariate matrix.

# Propose: Weighted PPLS

- Weighted PPLS
  - A weighted PLS model with a conjugate gradient path.
  - Penalized regression in the framework of weighted PLS.
  - Similar to the PPLS construction.
- Weighted PLS model:  $Y = b_0 + \sum_{i=1}^p b_i (X_i - \bar{x}_i \mathbf{1})$
- Penalize  $b_i$  by soft-thresholding:

$$b'_i = \arg \min_{\beta_i} (\beta_i - b_i)^2 + \lambda |\beta_i|$$

- $b'_i = \text{sign}(b_i) (|b_i| - \lambda)_+$ .
- $f_+ = \max(f, 0)$ .
- $\lambda$ : shrinkage parameter.

# Experiment: Combined data

- LOOCV error of predicting Minnesota samples
  - Weighted PPLS classifiers.
  - Top 200 genes

$w = PGA$	Shrink 0%					Shrink 40%					Shrink 80%				
	PLS components $k$					PLS components $k$					PLS components $k$				
	1	2	3	4	5	1	2	3	4	5	1	2	3	4	5
0	5	9	6	7	7	5	8	8	6	5	5	8	7	6	7
1/4	6	8	6	6	5	5	7	4	3	5	8	7	2	4	3
1/2	5	8	6	4	6	6	7	3	4	5	7	7	2	3	2
3/4	4	7	7	4	6	6	7	4	4	6	8	6	2	2	2
1	5	7	7	4	7	6	7	4	4	6	8	5	3	2	2

# Summary

- Weighted PPLS:
  - Penalized regression in a framework of weighted PLS.
  - Penalization/shrinking can improve over weighted PLS.
- Weighted PPLS methods with combined data:
  - Account for possible different relevances of the other studies by weighting.
  - Improve the performance of the classifier using data from a single study.

# General application

- Broad scope of the weighting scheme:
  - Applicable when the PGA data only contain ischemic and normal groups.
- Further extendable:
  - The primary and secondary experiments were conducted under different (but relevant) conditions, or on different organisms.
  - Microarray data with a survival end point.
  - Other loss functions.

# Reference

- **Huang, X., Pan, W., Grindle, S., Han, X., Chen, Y., Park, S.J., Miller, W.L., Hall, J. (2005).** A comparative study of discriminating human heart failure etiology using gene expression profiles. *BMC Bioinformatics* 6:205.
- **Huang, X., Pan, W., Han, X., Chen, Y., Miller, W.L., Hall, J. (2005).** Borrowing information from relevant microarray studies for sample classification using weighted partial least squares. *Computational Biology and Chemistry* 29(3), 204-211.